

ISSN 0974-3618 (Print)
0974-360X (Online)

www.rjptonline.org



REVIEW ARTICLE

Role of Myokines and prospects for their role in Diabetes Mellitus Therapy

**Kharissova Nuriya¹, Mindubayeva Farida^{1*}, Rajkamal Sharma³, Smirnova Liliya²,
Mkhitaryan Xeniya¹, Chergizova Bibigul¹, Salikhova Yelena^{1*}, Niyazova Yuliya¹,
Ryspayeva Gulnur¹, Evnevich Anna¹, Akimzhanova Neylya¹, Sarsembayeva Sholpan¹**

¹Karaganda Medical University” NCJSC, Karaganda, Kazakhstan.

²Kostroma State University, Kostroma, Russia.

³Advisor Cum Administrator Dr. Sarvesh Shukla Group Of Institution, Jaipur, Rajasthan, India.

*Corresponding Author E-mail: **7554422@mail.ru, Salehova_89@mail.ru**

ABSTRACT:

In the last decade, the attention of researchers has been drawn to the ability of skeletal muscles to produce biologically active substances (myokines). To date, several hundred myokines have been identified in the muscle secretome. Myokines have autocrine and paracrine effects. They have their own receptors in various tissues and organs. At certain concentrations, myokines have a systemic effect on tissues and organs, provide metabolic interaction between them and have a huge range of physiological effects. However, the biological activity of many of these myokines and their mechanism of action are either not yet characterized or poorly understood. Modern research is aimed at developing drugs that block myokine signaling pathways and studying the possibilities of their use in the treatment of neuromuscular diseases, obesity, type 2 diabetes mellitus, orthopedic pathology, as well as a decrease in muscle mass and muscle strength. Type 2 diabetes mellitus (T2DM) is a socially significant disease. Currently, there is no effective therapy to completely eradicate/cure diabetes and its associated complications. It is now necessary to consider in more detail the molecular pathways and targets for each pharmacological drug. There is a need to create new anti-diabetic therapy in the future based on myokines, knowing their signaling pathways and their mechanism of action on target cells, but also for the best possible combination therapy and strategies using available drugs and the beneficial effects of physical activity and exercise in the prevention and treatment of T2DM. A few studies in mice and humans have shown that exercise increases the levels of numerous myokines in the blood plasma, leading to the process of active transcription of myokines and accelerating metabolic processes associated with increased load on muscle tissue. It has been suggested that the secretion of myokines depends on the degree of physical training; intensity and duration of the athlete's training; its physiological and anatomical structure; the sport in which the athlete plays. Further scientific research will provide the key to understanding the process of secretion of myokines (proteins) in the body and the mechanism of their effect on various organs/systems and tissues, which will undoubtedly contribute to the success of doctors in the field of practical healthcare in the correction of pathological disorders, including diabetes mellitus.

KEYWORDS: Myokines, Muscles, Physical Exercise, diabetes mellitus type 2, metabolic disease.

INTRODUCTION:

1. Myokines, its function and mechanisms of action on various organs and tissues:

Recently, attention has been paid to the study of the endocrine function of muscle tissue - the secretion of myokines. The formation of myokines consists of two stages: transcription, that is, the rewriting of information from DNA genes to messenger RNA, and translation,

Received on 15.02.2024 Modified on 06.05.2024
Accepted on 27.06.2024 © RJPT All right reserved
Research J. Pharm. and Tech 2024; 17(10):5119-5131.
DOI: 10.52711/0974-360X.2024.00786

the direct assembly of myokine molecules^{1,2}. Myokines are cytokines, peptides, and proteoglycans produced, expressed, and released by muscle fibers (myocytes) in response to muscle contraction that have autocrine, paracrine, or endocrine effects in regulating metabolism in other tissues^{3,4}. The influence of myokines occurs through signaling pathways involved in the pathogenesis of many socially significant diseases, usually affect older people: abdominal obesity, type 2 diabetes mellitus (DM), cardiovascular and neurodegenerative pathologies, colon cancer and breast cancer^{5,6}. Pedersen et al. put forward the idea of “morbidoma” (originally “diseasome”) - a nosological cluster based on the persistence of chronic low-active inflammation, which is a common feature of the pathogenesis of diabetes mellitus, obesity, atherosclerosis, neurodegenerative lesions⁷. The experiment recorded an exponential increase in the concentration of interleukin in proportion to the duration of the contraction and the amount of muscle mass involved in it⁸. The first muscle-derived myokine identified was myostatin⁹. Currently, more than 1000 myokines are known, belonging to various structural and functional groups (cytokines, chemokines, prostaglandin family, etc.)¹⁰.

Due to the significantly increased number of molecules that have been identified as myokines, many authors have addressed the problem of their classification, according to chemical nature and physiological effect. Silja Raschke and Jurgen Eckel¹² proposed to form three groups - myokines, adipomyokines and adipokines^{11,12,13,14,15}. The discovery of myokines and the mechanisms of their secretion into the blood is the basis for uncovering the mechanisms of the influence of physical activity on metabolism and anti-inflammatory effects. Physical activity of varying intensity leads to the launch of a large number of biochemical, molecular and genetic mechanisms that underlie the body's adaptive reactions to physiological stress^{15,16,17,18}. Myokines perform a variety of functions during physical activity^{19,20}. As a consequence, myokines are considered important factors in maintaining homeostasis and muscle adaptation to physical activity²¹. The discovery of myokines has opened a new direction in understanding the effects of exercise on humans²².

Myokines play a role in the implementation of numerous processes, such as myogenesis, osteogenesis, thermogenesis, lipolysis, increasing the sensitivity of tissues to glucose^{23,24,25}. Type 2 diabetes mellitus (T2DM) is a socially significant disease. Insulin resistance (IR) is the first link in the pathogenesis of T2DM and is associated with a decrease in the ability of insulin to enhance the uptake of glucose from the bloodstream by target cells. The molecular mechanisms and markers of IR have not been sufficiently studied,

which hinders early diagnosis and the development of preventive drug therapy. Reliable global estimates of diabetes prevalence are needed to monitor public health strategies and develop effective future interventions. Treatment of diabetes is aimed at preventing complications and maintaining a satisfactory quality of life. Many drugs are currently being developed to treat diabetes.^{26,27,28,29,30}

Myokines are formed, expressed on the surface of muscle fibers and released into the bloodstream. They are regulators of myogenic differentiation, fiber type switching, and muscle mass maintenance^{31,32}. Myokines are involved in the regulation of various physiological processes and have both local and systemic effects³³. Myokines are involved in bone metabolism, influencing bone resorption and formation by interacting with factors associated with bone cell secretion or influencing bone metabolic pathways³⁴.

Experiments on mice showed that after physical exercise there was an increase in the concentration of osteocalcin (OCC) and the myokine IL-6, but in the presence of IL-6 deficiency, the level of OCC did not change³⁵.

It is also worth noting that during physical activity the level of myostatin decreases, thereby stimulating the process of muscle tissue growth^{36,37,38}. Myokines play a key role in regulating myocardial and skeletal muscle rejuvenation, weakening muscle metabolic homeostasis, and protecting against ischemic damage and apoptosis³⁹.

Fibroblast growth factor (FGF-21) improves tissue sensitivity to insulin²⁴. Myostatin inhibits the growth and differentiation of muscle tissue and regulates the growth of adipocytes⁴⁰. Myostatin leads to resorption in bone tissue, while increasing osteoclastogenesis and inhibiting osteoblastogenesis⁴¹. Irisin improves lipid metabolism, facilitates the deposition of glucose in the liver, and prevents the occurrence of hyperglycemia and hyperlipidemia. It has a beneficial effect in the pathogenesis of obesity and type 2 diabetes^{24,42}. Interleukin-6 (IL-6) promotes the development of insulin resistance (IR) in adipose tissue and hepatocytes during physical exercise (PE) for more efficient mobilization of glucose and fatty acids as energy sources, as well as in obesity, metabolic syndrome, diabetes mellitus^{43,44}. IL-6 in the blood and tissues serves as a signal of energy deficiency, which enhances the action of insulin in muscle cells and inhibits it in tissues supplying energy substances. Its increased level leads to a lipolytic effect^{23,24,45}. IL-6 enhances lipolysis and insulin sensitivity in adipose tissue⁴⁶, optimizes insulin production in the pancreas⁴⁷ and increases glycogenolysis and lipolysis in the liver⁴⁸. A sharp increase in plasma concentrations of IL-6 has an anti-

inflammatory effect and regulates acute inflammatory reactions⁴⁹.

Modern studies demonstrate the active participation of myokines in the regulation of the processes of lipolysis, gluconeogenesis, insulin secretion by pancreatic beta cells, and activation of thermogenesis^{50,51,52}. Myokines are capable of modulating the function of cells of the immune system, local blood flow, energy metabolism, processes of proliferation and differentiation of myoblasts^{53,54,55,56}. Adipokines and myokines can serve as indicators of obesity phenotypes. Significant differences in the levels of adipokines and myokines and their relationship with indicators of the intestinal microbiome were revealed in patients with different obesity phenotypes^{57,58}. Changes in myokine production in diabetes mellitus have been shown^{59,60,61,62}, obesity^{23,63,64} osteoporosis^{65,66} as well as for metabolic disorders⁶⁷.

It is known that a sedentary lifestyle is associated with the development of obesity, type 2 diabetes, cardiovascular disease, osteoporosis and early mortality, and regular physical activity helps prevent these conditions⁶⁸. By adapting to mechanical, neural and humoral influences, skeletal muscle plays a critical role in physical activity, energy expenditure and glucose utilization⁶⁹. Many research projects led to the discovery of new myokines and an understanding of their role in the physiological regulation of exercise, both in apparently healthy people and in chronically suffering patients⁷⁰. During physical exercise, the following reactions occur in skeletal muscle: an increase in the expression of insulin receptors, the absorption of fatty acids and glucose, glycogen stores, as well as mitochondrial biogenesis. In a study of muscle secretions, hundreds of substances were found to be expressive in response to exercise⁷¹. Loss of muscle mass in people with chronic heart disease increases risk of death⁷². During exercise, muscle fiber cells express and release more than 3,000 molecules of cytokines and polypeptides known as “myokines”⁷³. Myokines have a positive effect on the development of satellite cells and skeletal muscle hypertrophy⁷⁴. Myokines interact with high-affinity receptors (transmembrane glycoproteins) located on the membrane of target cells⁷⁵. After exercise, there is an increase in PGC-1 α (Peroxisome proliferator-activated receptor gamma coactivator 1-alpha), which stimulates mitochondrial biogenesis, binds to FOXO and inhibits its transcriptional activity, thereby preventing the breakdown of muscle proteins⁷⁶. The primary role of IL-6 is to maintain stable blood glucose levels during exercise. Low pre-exercise levels of intramuscular glycogen have been shown to increase mRNA synthesis IL-6⁷⁷. Oxidative stress, low glucose concentration, low glycogen content and increased Ca²⁺ flux activate IL-6

transcription factors by stimulating mitogen-activated protein kinases⁷⁸. Skeletal muscle secretes myokines, while influencing other organs by binding to target cell receptors in an autocrine, endocrine and paracrine manner⁷⁹.

2. The role of myokines in diabetes mellitus:

Myokines promote increased insulin sensitivity, which leads to glucose clearance and regulation of glucose and lipid metabolism⁸⁰. However, long-term hyperinsulinemia leads to dysfunction of pancreatic β -cells and decreased insulin synthesis against the background of persistent IR. Prediabetes occurs, in which carbohydrate metabolism is disrupted. At this stage, it is possible to prevent pathological changes and restore the sensitivity of target cells to insulin. In this regard, early diagnosis of IR is of great practical importance^{81,82}. The development of insulin resistance leads to the inability of target cells to respond to insulin, resulting in the development of type 2 diabetes mellitus (T2DM) and metabolic syndrome⁸³. Kinases such as NF- κ B, IKK, JNK are involved in inflammatory signaling cascades in adipocytes under the influence of cytokines⁸⁴. IRS is used as one of the substrates, which is phosphorylated at several serine residues, rendering it inactive and interrupting signal transmission along the insulin cascade^{85,86,87,88,89}. Cytokines and free fatty acids (FFA) have a pro-inflammatory effect. Cytokines and FFAs bind to toll-like receptors TLR4 on adipocytes. An inflammatory cascade is launched involving NF- κ B, IKK and JNK. As a result, serine phosphorylation of IRS increases in adipocytes and the insulin cascade is switched off^{90,91}.

Many antihyperglycemic drugs used to treat diabetes (thiazolidinediones, sulfonylureas, glinides, insulin) activate the key regulator of adipogenic differentiation PPAR γ , causing weight gain due to the formation of new fat depots⁹². Studies of cell insulin cascades have shown the possibility of selective effects on cell growth and division in the absence of induction of lipo- and adipogenesis. The transcription factor Prepl of the TALE family of homeobox proteins not only regulates PPAR γ activity, but also enhances the expression of the glucose transporter Glut4 and insulin sensitivity^{93,94,95}.

In pharmacological therapy of T2DM, differentiation of adipose tissue preadipocytes into brown and beige fat cells, which have hypolipidemic and hypoglycemic properties, can be used. Their cells promote the utilization of fats and glucose without obtaining energy. Its key participant is the protein UCP-1, or thermogenin. The expression of thermogenin and the differentiation of fat progenitor cells into “beige” adipocytes is controlled by the local hormone irisin⁹⁶⁻⁹⁷.

It has been noted that current antidiabetic drugs can cause serious side effects, including heart failure, hepatotoxicity and obesity. Scientists' work aimed at elucidating the mechanism of skeletal muscle sensitivity to insulin, energy metabolism and the role of myokines in endocrine functions will help find new treatment methods in the future⁹⁸. Effect of myokines secreted from healthy and insulin-resistant myotubes is beneficial and harmful in the regulation of insulin secretion, which indicates cross-talk between skeletal muscles and the pancreas⁹⁹. As expected, there has been an uptick in research into contraction-regulating myokines that promote β -cell mass and function^{100,101}. Beneficial myokines, including irisin, fractalkine, FGF21, myonectin, and IL-15, improve the mass and/or function of β -cells that regulate glucose and lipid metabolism^{102,103}.

Myostatin can be used to prevent muscle wasting¹⁰⁴. An example of disparate results is shown in the myokine IL-6, where some investigators report that IL-6 has beneficial effects on β -cell function^{105,106,107,108}, which casts doubt on the appropriateness of using irisin as a therapeutic agent¹⁰⁸. Basically, experimental studies have been conducted in rodent models to identify the mechanism of action of myokines on the viability and function of β -cells^{109,110}.

Persistent IR triggers delayed, long-term adaptation processes in the body; they lead to a restructuring of the entire metabolism during the development of T2DM. Such long-term changes are consolidated at the transcriptional level, where various transcription factors play a key role¹¹¹.

In an experiment on mice that were fed a high-fat diet and periodic chronic stress, glucose homeostasis and lipolysis were determined. In the early stages of diet-induced obesity, it was found that glucose intolerance initially occurs, followed by decreased insulin sensitivity and increased sensitivity to epinephrine (EPI). Stress increased myonectin (Myn) levels in serum and skeletal muscle and improved glucose intolerance^{112,113}.

Decreased insulin receptor sensitivity worsens insulin effects, triggering a vicious cycle of disorders including compensatory insulin hypersecretion, pancreatic β -cell dysfunction, impaired tissue glucose utilization, metabolic and hemodynamic abnormalities, subsequent increases in IR, organ damage, and cardiovascular disease (CVD). Early therapy aimed at improving insulin sensitivity and restoring its effects could become one of the effective methods of primary prevention of type 2 diabetes and CVD¹¹⁴.

A study conducted on overweight and obese Chinese residents found that low serum irisin levels are usually associated with smoking, type 2 diabetes mellitus, dyslipidemia, and the presence of cardiovascular disease risk factors¹¹⁵. The level of irisin concentration depends on the etiology of obesity. For example, with a genetic form of obesity in patients with Prader-Willi syndrome, the content of irisin in the blood serum is significantly lower than with other forms of obesity that are not caused by genetic factors¹¹⁶. In patients with type 2 diabetes mellitus, the level of irisin concentration in the blood serum is reduced¹¹⁷. Vascular complications are the cause of death in patients with type 2 diabetes; it was revealed that irisin prevents endothelial dysfunction by reducing oxidative stress through inhibition of signaling pathways including NF- κ B/iNOS and PKC-/NADPH oxidases¹¹⁸. Thus, irisin and its analogues may be potential drugs for the treatment of complications of diabetes mellitus¹¹⁹.

There are many models of spontaneous development of type 1 and type 2 diabetes in rodents (rats and mice). Diabetes can be caused by surgery, pharmacological drugs, the prescription of special diets, as well as various combinations of these¹²⁰.

The effect of forced running loads on the content of certain cytokines in the skeletal muscles of mice with a model of type II diabetes mellitus was studied. A high-fat diet was used to form a disease model; physical activity in the form of forced running was carried out for 4 weeks. The production of interleukins can be associated with the restructuring of transcriptional mechanisms in muscle fibers associated with calcium-dependent and calcium-independent intracellular signaling pathways^{121,123,124}.

The study of the role of myokines in the correction of various disorders has significant prospects. A change in the production of myokines has been shown in diabetes mellitus, obesity, osteoporosis, and also in metabolic disorders. This allows us to consider myokines as a possible mechanism through which the therapeutic effects of physical activity are realized in various diseases^{125,126}.

3. Prospects for myokines in the treatment of diabetes

Adipokines and myokines maintain metabolic homeostasis. When they are imbalanced, metabolic complications develop, such as insulin resistance, low-grade inflammatory remodeling of adipose tissue, extracellular lipid deposition, atherosclerosis, etc¹²⁷. Adiponectin has been shown to have insulin-sensitizing, antiatherogenic and anti-inflammatory effects¹²⁸. But the adipokines leptin and resistin, on the contrary, are

factors of insulin resistance. They have a pleiotropic effect on food intake, neuroendocrine regulation of the hypothalamus, reproductive function and energy metabolism¹²⁹. Data on the adipokine asprosin are extremely fragmentary: there are suggestions that it can influence glucose metabolism and is probably associated with obesity and that its level may be a marker for the early diagnosis of diabetes mellitus¹³⁰.

Information about the role of the myokine myostatin in the pathogenesis of obesity and insulin resistance is currently limited. Some authors propose the use of myostatin to identify metabolically unhealthy obesity (MHA)¹³¹. The insulin-sensitive myokine FGF21 acts as a metabolic regulator involved in the control of glucose homeostasis¹³². Recently, researchers have been interested in the peptide osteonectin, secreted by fibroblasts and osteoblast-like cells. Its pathophysiological role is poorly understood¹³³. Single results in patients with obesity and insulin resistance demonstrated a higher level compared to healthy subjects¹³⁴.

Exercise has long been a central principle of both the prevention and treatment of type 2 diabetes mellitus (T2DM)^{135,136}. The secretion of IL-6 during physical activity depends on the energy state of the cell and on the glycogen content in the cells before exercise. Low glycogen content leads to greater release of IL-6 under conditions of energy crisis in the muscle cell during contraction¹³⁷. In vitro studies demonstrate that IL-6 treatment increases glucose uptake through AMP protein kinase [adenosine monophosphate kinase (AMPK)] and phosphatidylinositol 3-kinase (PI3K) pathways¹³⁸.

Carey et al.¹³⁹ reported increased insulin-dependent glucose uptake in vivo in response to IL-6 infusion. In contrast, Harder-Lauridsen et al.¹⁴⁰ found no increase in glucose uptake during euglycemic hyperinsulinemic clamp with IL-6 infusion in individuals with T2DM, although there was a decrease in plasma insulin levels, indicating increased insulin sensitivity¹⁴⁰. Jiang et al.¹⁴¹ found a differential effect of IL-6 treatment on primary myotubes compared with normal glucose tolerance and T2DM, suggesting a minor role for IL-6 in T2DM muscle. IL-6 treatment enhanced both insulin-dependent and -independent glucose uptake and glycogen synthesis in healthy myotubes, but this effect was lost in T2DM myotubes. This suggests that from a glucose control perspective, the contraction-inducible myokine IL-6 is effective in preventing T2DM but may not be effective in glucose uptake in patients with existing T2DM. IL-13 is released from human primary myotubes in vitro and has been demonstrated to have an “insulin-like” effect on glucose metabolism in human muscle by increasing glucose uptake, glycogen synthesis, and glucose oxidation in primary myotubes in normal and T2DM¹⁴¹.

This “insulin-like” effect is mediated by activation of the Akt and PI3K pathways. IL-13 expression is increased in response to resistance training in human skeletal muscle¹⁴², but there is no evidence of increased plasma IL-13.

In addition, Gergens et al.¹⁴³ demonstrated the expression and release of FSTL-1 from primary human myotubes. Interestingly, contraction of primary ER myotubes did not induce FSTL-1 secretion; however, increases in circulating plasma FSTL-1 have been observed in humans following acute aerobic exercise. In vitro incubation of L6 myotubes (rat cell line) in FSTL-1 has been shown to increase glucose uptake in an AMPK- and calcium-calmodulin kinase-dependent manner¹⁴⁴, resulting in increased expression of GLUT4 mRNA and its translocation to the plasma membrane, mediating improved glucose control.

Optimizing myokine responses for the prevention and treatment of T2DM. Myokines that regulate glucose and fat metabolism can trigger mechanisms during exercise that prevent the onset or progression of T2DM¹⁴⁵. To date, most of the evidence describing the mechanism by which myokines modulate metabolic function has been characterized using in vitro cell models, which do not necessarily correspond to the in vivo human situation¹⁴⁶. Tingting Guo et al. investigated the effect of inhibition of myostatin signaling in skeletal muscle and adipose tissue on body composition, metabolic profile¹⁴⁷. Mice deficient in myostatin showed normal basal metabolic rate and higher respiratory quotient, indicating an increased rate of carbohydrate oxidation; an increase in the amount of lean mass and a low content of fat mass was also noted over 15 months of observation¹⁴⁸. Another study showed increased tissue sensitivity to insulin in mice deficient in myostatin due to increased AMPK activity in muscle¹⁴⁹. Skeletal muscle (SM) is known to generate hypoglycemic and antioxidant responses¹⁵⁰.

Myokines and exercise. Exercise is a well-established treatment for muscle metabolic defects that occur in T2DM.

Physical activity promotes the process of active transcription of myokines, which accelerate metabolic processes associated with increased stress on muscle tissue. Thus, by explaining the process of formation of these proteins in the body, doctors will be able to correct various disorders that are associated with the body's complex adaptation to physical activity after severe injuries^{151,152}. It is also worth noting that during physical activity the level of myostatin decreases, thereby stimulating the process of muscle tissue growth. Its concentration in mouse myocytes decreases after

running exercise, promoting the growth and differentiation of satellite cells¹⁵³. Similar results have been obtained in humans^{154,155}.

A 2020 meta-analysis including seven studies involving 125 normal-weight, overweight, and obese adults (21–64 years) found that bouts of physical activity (PE) increased serum FGF-21 levels regardless of body weight. In this case, the increased level of FGF-21 persists for 1 hour and decreases to a level close to the initial values after 3 hours¹⁵⁶. However, there was no increase in FGF-21 levels in patients with type 2 diabetes¹⁵⁷.

Studies in mice have shown that irisin improves glucose tolerance and reduces insulin resistance (IR)¹⁵⁸. Irisin also stimulates lipolysis using hormone sensitive lipase (HSL, hormone sensitive lipase) and inhibits lipogenesis in mouse adipocytes¹⁵⁹.

In the work of Miyamoto-Mikami E. et al. In healthy adults, after 8 weeks of endurance training, increases in circulating irisin levels were positively correlated with decreases in fat mass¹⁶⁰. Irisin has antioxidant and anti-inflammatory effects on hepatocytes, which could be useful in reducing the activity of steatohepatitis¹⁶¹.

Some of the best studied exercise-induced myokines influence bone formation and bone resorption¹⁶².

Dynamic and static exercises differentially affect the content of myokines in the blood plasma of athletes and untrained individuals^{24,156,160,163}.

A study of the effect of forced running in mice with a model of type II diabetes mellitus showed that in healthy animals there was a decrease in the concentrations of IL-6 and IL-15 and an increase in the concentration of leukemia inhibitory factor (LIF) in muscle tissue after 4 weeks of regular forced running. At the same time, in diabetic mice, the concentrations of IL-6 and IL-15 after exercise increased, and LIF, on the contrary, decreased¹⁶⁴.

The production of interleukins can be associated with the restructuring of transcriptional mechanisms in muscle fibers associated with calcium-dependent and calcium-independent intracellular signaling pathways^{165,166,167,168}.

An increase in IL66 mRNA transcription was shown in the nuclei of muscle cells isolated from human muscle biopsies after a single exercise^{169,170}.

This is also of interest given that lack of physical activity is strongly associated with the occurrence of

T2DM. There is a need to determine the secretion of myokines in patients with T2DM to identify potential therapeutic targets for the treatment of this disease¹⁷¹. The synthesis of IL-6 during physical exercise is due to glycogen-independent mechanisms and changes in the level of calcium in the cytosol of myocytes. It consists primarily of regulating glucose homeostasis, which is necessary for intensely contracting muscles. With an increase in the intensity of muscle contractions and a decrease in glycogen concentration in the muscles, mechanisms associated with the activation of mitogen-activated protein kinase are activated¹⁷².

4.Challenges and limitations associated with the use of myokines for therapeutic purposes:

The occurrence of diabetes mellitus is associated with myokines, but the mechanism of damage to target organs is still unclear and requires further study. Cytokines such as interleukin-4 and interleukin-10 are capable of suppressing type 1 diabetes in animal models when delivered as part of a plasmid or viral vector¹⁷³.

When creating a new drug, it takes a long time from the idea to its entry into the market and widespread use in practice. The algorithm for entering the market of a new pharmaceutical has the following stages: 1) idea, 2) development of production technology, 3) preclinical testing of the pharmaceutical (previously only on animals, now also on cell cultures in in vitro models), 4) after clinical trials (CT), 5) entering the market, 6) additional clinical trials (CT) are being conducted at a new level¹⁷⁴.

Experts from Tomsk State University (TSU) are developing a new method of treating diabetes¹⁷⁵.

Although an increase in the number of clinical trials indicates renewed interest in cytokines, antibodies targeting cytokines or corresponding receptors are being used more often¹⁷⁶.

Exercise-induced increases in IL-6 production may be involved in protecting pancreatic β -cell mass and function. It is well known that exercise can improve insulin sensitivity, whereas it is less clear whether exercise can improve insulin secretion and whether there is a link between insulin-resistant skeletal muscle and pancreatic β -cells^{172,177,178}.

In diabetes mellitus, glucose metabolism is impaired, which leads to increased blood glucose levels (hyperglycemia) and the production of free radicals. Drugs, used to treat metabolic disorders are not flawless. Basic knowledge of diabetic patients regarding insulin therapy is insufficient. Urgent steps are needed to improve the knowledge of diabetic patients and create

appropriate awareness about insulin therapy. Complications of diabetes can be avoided by regularly monitoring and maintaining blood glucose levels within normal limits. The current approach to treating both type 1 and type 2 diabetes is to achieve the best possible glucose control^{179,180,181,182,183}. In diabetes, self-monitoring and patient compliance with prescribed medications, as well as lifestyle changes, are very important, and the pharmacist can play an important role in counseling¹⁸⁴.

A promising direction is the introduction of artificial intelligence in the management of diabetes. Artificial intelligence is able to correctly interpret external data, learn from such data and use the acquired knowledge to achieve specific goals and objectives through flexible adaptation¹⁸⁵. The artificial intelligence of the SNAQ app automatically identifies foods and calculates the protein, fat and carbohydrate content. Throughout the day, the app receives self-monitoring data and marks meal times on a graph, helping to illustrate how certain foods affected your blood glucose levels. Using the SNAQ app as part of nutrition counseling may be a promising technology as it provides real-time information on food intake and allows for personalized recommendations^{186,187}.

Currently, there is no effective therapy to completely eradicate/cure diabetes and its associated complications. There is a need to create better, new anti-diabetic therapy in the future based on myokines, knowing their signaling pathways and their mechanism of action on target cells, but also for the best possible combination therapy and strategies using available drugs¹⁸⁸.

Neuromuscular electrical stimulation (NMES) is emerging as an effective exercise substitute for myokine induction. NMES is safe and effective and has been shown to improve muscle strength, functionality and quality of life. This alternative method of exercise induces hypertrophy and neuromuscular adaptations in skeletal muscles. NMES stimulates the secretion of circulating myokines, promoting a cascade of endocrine, paracrine and autocrine effects. NMES is an effective exercise replacement for stimulating myokine production and its potential applications in health and disease¹⁸⁹.

5. Key findings and insights regarding the role of myokines in diabetes mellitus:

Diabetes mellitus (DM) has become a global threat to human health around the world. Currently, there is no effective therapy to completely eradicate/cure diabetes and its associated complications. Current antidiabetic drugs (thiazolidinediones, sulfonylureas, glinides, insulin) can cause serious side effects, including heart failure, hepatotoxicity, and obesity.

The work of scientists aimed at elucidating the mechanism of skeletal muscle sensitivity to insulin, energy metabolism and the role of myokines in endocrine functions will help in the future to find new treatment methods based on knowledge of signaling pathways and the mechanisms of their action on target cells.

They found that myokines promote insulin sensitivity, which leads to glucose clearance and regulation of glucose and lipid metabolism. Several myokines (irisin, fractalkine, FGF21, myonectin, and IL-15) improve the mass and/or function of β -cells, which regulate glucose and lipid metabolism.

Myokines as therapeutic targets for the treatment of T2DM are a promising area. Harnessing the beneficial effects of myokines on insulin secretion, insulin sensitivity, and energy metabolism may lead to the development of important new treatments for T2DM.

A few studies in mice and humans have shown that exercise increases the levels of numerous myokines in the blood plasma, which leads to the process of active transcription of myokines and acceleration of metabolic processes associated with increased load on muscle tissue.

It has been suggested that the secretion of myokines depends on the degree of physical fitness; intensity and duration of the athlete's training; its physiological and anatomical structure; the sport in which the athlete plays. Neuromuscular electrical stimulation (NMES) is emerging as an effective replacement for myokine induction by exercise.

A promising direction is the introduction of artificial intelligence in diabetes management.

CONFLICT OF INTEREST:

The authors declare that there is no conflict of interest.

REFERENCES:

1. Tsoriev T.T., White Zh.E., Rozhinskaya L.Ya. The role of myokines interstitial interaction and regulation of metabolism: a review of literature. Osteoporosis and Bone Diseases. 2016; 19(1): 28-34. (In Russ.) <https://doi.org/10.14341/osteo2016128-34>
2. Pedersen B.K., Febbraio M.A.. Muscle as an Endocrine Organ: Focus on Muscle-Derived Interleukin-6. *Physiol Rev.* 2008; 88(4): 1379-1406. <https://doi.org/10.1152/physrev.90100.2007>
3. Pedersen BK, Åkerström TC, Nielsen AR, et al. Role of myokines in exercise and metabolism. *J Appl Physiol.* 2007; Sep; 103(3): 1093-1098. doi: 10.1152/japplphysiol.00080.2007. Epub 2007 Mar 8
4. Grebennikova TA, Belaya ZhE, Tsoriev TT, et al. Endocrine function of bone tissue. Osteoporoz i osteopatii. 2015; (1): 28-37. (In Russ)
5. L Gameau, C Aguer Role of myokines in the development of skeletal muscle insulin resistance and related metabolic defects in

- type 2 diabetes *Diabetes Metab.* 2019; Dec; 45(6): 505-516. DOI:10.1016/j.diabet.2019.02.006. Mar4. PMID: 30844447 DOI: 10.1016/j.diabet.2019.02.00645(6):505-516. doi: 10.1016/j.diabet.2019.02.006. Epub
6. Jenny Hyosun Kwon, Kyoung Min Moon, and Kyueng-Whan Min Exercise-Induced Myokines can Explain the Importance of Physical Activity in the Elderly: An Overview *Healthcare* 2020; 8(4): 378; <https://doi.org/10.3390/healthcare8040378>
7. Pedersen B.K. Muscle as a secretory organ. *Compr. Physiol.* 2013; 3: 1337-1362.
8. van Hall G, Steensberg A, Sacchetti M, et al. Interleukin-6 Stimulates Lipolysis and Fat Oxidation in Humans. *J Clin Endocrinol Metab.* 2013. Chowdhury S, Schulz L, Palmisano B, et al. Muscle-derived interleukin 6
9. Allen D.L., Cleary A.S., Speaker K.J et al. Myostatin, activin receptor IIb, and follistatin-like-3 gene expression are altered in adipose tissue and skeletal muscle of obese mice. *Am. J. Physiol. Endocrinol. Metab.* 2008 V. 294 P. 918-927
10. Exercise-Induced Myokines can Explain the Importance of Physical Activity in the Elderly: An Overview by Jenny Hyosun Kwon 1,Kyoung Min Moon 2, and Kyueng-Whan Min *Healthcare* 2020; 8(4): 378; <https://doi.org/10.3390/healthcare8040378>
11. Orlov S.N., Kapilevich L.V., Dyakova E.Yu., Zakharova A.N., Kabachkova A.V., Kalinnikova Yu.G., Klimanova E.A., Kironenko T.A., Milovanova K.V., Sidorenko S.V. Skeletal muscles as an endocrine organ. – Tomsk: Publishing House of Tomsk State University, 2018. – 190 p. ISBN 978-5-94621-765-1.
12. Raschke S., Eckel J. Adipo-myokines: two sides of the same coin-mediators of inflammation and mediators of exercise // *Mediators Inflamm.* 2013 (2013): 320724
13. Simbirtsev A.S. Cytokines: classification and biological functions // *Cytokines and inflammation.* 2004; No. 3 P. 16-22;
14. Borish L.C., Steinke J.W. Cytokines and chemokines // *J. Allergy Clin. Immunol.* 2003 V. 111, No. 2 P. 460-475; Lata S., Raghava G.P.S. CytoPred: a server for prediction and classification of cytokines // *Protein Eng. Des. Sel.* 2008 V. 21, No. 4 P. 279-282.
15. Rian Q Landers-Ramos I, Nathan T Jenkins, Espen E Spangenburg, James M Hagberg, Steven J Prior Circulating angiogenic and inflammatory cytokine responses to acute aerobic exercise in trained and sedentary young men PMID: 24643426 PMID: PMC4048778 DOI: 10.1007/s00421-014-2861-6.
16. Kharisova N., Smirnova L., Kuzmin A., et al. The influence of the physical activity of a modern student on the characteristic of the CVS and RS and their resistance to stress during educational process. *Georgian Medical News.* 2019; 12 (297) ; 124-129. PMID: 32011307
17. Hittel DS, Axelsson M, Sarna N, et al. Myostatin Decreases with Aerobic Exercise and Associates with Insulin Resistance. *Med Sci Sport Exerc.* 2010; 42(11): 2023-2029. <https://doi.org/10.1249/MSS.0b013e3181e0b9a8>.
18. Ryan AS, Li G, Blumenthal JB, Ortmeier HK. Aerobic exercise + weight loss decreases skeletal muscle myostatin expression and improves insulin sensitivity in older adults. *Obesity.* 2013; 21(7): 1350-1356. <https://doi.org/10.1002/oby.20216>
19. Rudnick J., Püttmann B., Tesch P.A. et al. Differential expression of nitric oxide synthases (NOS 1-3) in human skeletal muscle following exercise countermeasure during 12 weeks of bed rest // *FASEB J.* 2004 V. 18, No. 11 P. 1228-1230
20. Otis J.S., Burkholder T.J., Pavlath G.K. Stretch-induced myoblast proliferation is dependent on the COX2 pathway // *Exp. Cell Res.* 2005 V. 310 P. 417-425.
21. Broholm C., Pedersen B.K. Leukaemia inhibitory factor – An exercise-induced myokine // *Exercise Immunology Review.* 2010 V. 16 P. 77-85.
22. Barbalho, Sandra and Neto, Edmundo and Goulart, Ricardo and Bechara, Marcelo and Chagas, Eduardo and Audi, Mauro and Campos, Leila and Guiger, Elen and Buchaim, Rogério and Buchain, Daniela and Cressoni Araujo, Adriano. (2020). Myokines: A descriptive review. *The Journal of sports medicine and physical fitness.* 60. 10.23736/S0022-4707.20.10884-3.
23. Pedersen B.K.; Febbraio, M.A. Muscles, exercise and obesity: Skeletal muscle as a secretory organ. *Nat. Rev. Endocrinol.* 2012, 8, 457-465.
24. Mai Charlotte Krogh Severinsen, Bente Klarlund Pedersen Muscle-Organ Crosstalk: The Emerging Roles of Myokines. *Endocr Rev.* 2020; Aug 1; 41(4): 594-609. PMID: 32393961. PMID: PMC7288608. doi: 10.1210/endrev/bnaa016.
25. Sorokina L. D., Marchenko E. A., Zavyalova, A. N. Myokines. Literature review. *force*, 5(S3), 947-948. (In Russ.) Volume 5 No.. S3 (2022): Materials of the All-Russian scientific forum of students with international participation “Student Science - 2022” / Miokins. Literature review <https://ojs3.gpmu.org/index.php/forcipe/article/view/5438>
26. Bhutkar M. A., Bhise S. B. Comparative Studies on Antioxidant Activity of Some Antidiabetic Plants. *Research J. Pharm. and Tech.* 2011; 4(9): Sept. 1409-1412.
27. Hepcy Kalarani D, Venkatesh P, Dinakar A. Anti-Diabetic Activity of Aqueous Extract of Leaves of Pavonia zeylanica in Rats. *Research J. Pharm. and Tech.* 2009; 2(4): 789-792.
28. P.M. Patil, P.D. Chaudhari, N.J. Duragkar, P.P. Katolkar. Formulation and Evaluation its Anti-diabetic Activity of Liquid Oral Preparation of Gymnema sylvestre and Stevia rebaudiana and their Combination in Alloxan Diabetic Rats. *Research J. Pharm. and Tech.* 2010; 3(4): 1200-1204.
29. Sandeep Goyal, V.K. Bansal, Dhruva Sankar Goswami, Suresh Kumar. sVascular Endothelial Dysfunction: Complication of Diabetic Mellitus and Hyperhomocysteinemia. *Research J. Pharm. and Tech.* 2010; 3(3): 657-664.
30. M Yashpal Naidu, K P Channa Basavaraj, T Tamizh Mani, K Roopa. Validated RP-HPLC Method for the Quantitation of Pioglitazone an Anti - Diabetic Drug in Bulk and Pharmaceutical Dosage Forms. *Research J. Pharm. and Tech.* 2010; 3(3): 885-887.
31. Paltsyn A A Myokines Pathological physiology and experimental therapy. 2020; 64(1): 135-141 DOI: <https://doi.org/10.25557/0031-2991.2020.01.135-141>
32. Wentao Chen, Liyi Wang, Wenjing You, Tizhong Shan Myokines mediate the cross talk between skeletal muscle and other organs. *J. Cellular physiology.* 2021; 236(4) April; 2393-2412. <https://doi.org/10.1002/jcp.30033> Chen W, Wang L, You W, Shan T. Myokines mediate the cross talk between skeletal muscle and other organs. *J Cell Physiol.* 2021; Apr; 236(4): 2393-2412. doi: 10.1002/jcp.30033. Epub 2020 Sep 3. PMID: 32885426.
33. Moonesan MR. Muscle-kidney crosstalk; the role of myokines. *J Ren Endocrinol.* 2023; 9: e25129. doi: 10.34172/jre.2023.25129. *J Ren Endocrinol* 2023; 9: e25129. Nickan Research Institute Journal of Renal Endocrinology <https://www.jrenendo.com> doi: 10.34172/jre.2023.25129
34. QiYang Wang, QiuNan Lv, YuQiong Zhang, GuoXi Gao, Sheng Lu Advances in the research on myokine-driven regulation of bone metabolism Ming Hong Shao, 2023; DOI: <https://doi.org/10.1016/j.heliyon.2023.e22547>
35. Chowdhury S, Schulz L, Palmisano B, et al. Muscle-derived interleukin 6 increases exercise capacity by signaling in osteoblasts. *J Clin Invest.* 2020; 130(6): 2888-2902. <https://doi.org/10.1172/JCI133572>
36. Matsakas A, Friedel A, Hertrampf T, Diel P. Short-term endurance training results in a muscle-specific decrease of myostatin mRNA content in the rat. *Acta Physiol Scand.* 2005; 183(3): 299-307. <https://doi.org/10.1111/j.1365-201X.2005.01406.x>
37. Kainulainen H, Papaioannou KG, Silvennoinen M, et al. Myostatin/ activin blocking combined with exercise reconditions skeletal muscle expression profile of mdx mice. *Mol Cell Endocrinol.* 2015; 399: 131-142. <https://doi.org/10.1016/j.mce.2014.10.001>
38. Ko IG, Jeong JW, Kim YH, et al. Aerobic Exercise Affects Myostatin Expression in Aged Rat Skeletal Muscles: A Possibility of Antiaging Effects of Aerobic Exercise Related With Pelvic Floor Muscle and Urethral Rhabdosphincter. *Int Neurourol J.* 2014; 18(2): 77. <https://doi.org/10.5213/inj.2014.18.2.77>

39. Berezin AE, Berezin AA, Lichtenauer M. Myokines and Heart Failure: Challenging Role in Adverse Cardiac Remodeling, Myopathy, and Clinical Outcomes. *Dis Markers*. 2021;13:2021:6644631. PMID: 33520013. PMCID: PMC7819753.
40. Li F, Yang H, Duan Y, Yin Y. Myostatin regulates preadipocyte differentiation and lipid metabolism of adipocyte via ERK1/2. *Cell Biol Int*. 2011; 35(11): 1141-1146. <https://doi.org/10.1042/CBI20110112>
41. Qin Y, Peng Y, Zhao W, et al. Myostatin inhibits osteoblastic differentiation by suppressing osteocyte-derived exosomal microRNA-218: A novel mechanism in muscle-bone communication. *J Biol Chem*. 2017; 292(26): 11021-11033. <https://doi.org/10.1074/jbc.M116.770941>
42. Inyushkin A.N., Isakova T.S., Inyushkin A.A., Kretova I.G.. Physiological and pathophysiological role of the myokine irisin. *Modern issues of biomedicine*. 2023; 7(2) DOI: 10.51871/2588-0500_2023_07_02_8 UDC 612.018.2
43. Parakhonsky A.P. The role of interleukin-6 in the development of insulin resistance. *Advances in modern science*. 2011.No.1.:105-106; (In Russ. URL: <https://natural-sciences.ru/ru/article/view?id=15709>
44. Karimov R.N., Omonov M.O., Kamilova S.A., Abdullaev S.O. The role of interleukin-6 (IL-6) in the pathogenesis of diabetes type 2 mellitus. Volume-22_Issue-3_February_2023 *Journal of New Century Innovations*. <http://www.newjournal.org/>
45. Vasyukova O.V., Kasyanova Yu.V., Okorokov P.L., Bezlepina O.B. Myokines and adipomyokines: inflammatory mediators or unique molecules of targeted therapy for obesity? *Problems of Endocrinology*. 2021; 67(4): 36-45. (In Russ.) <https://doi.org/10.14341/probl12779>
46. Lutosławska G. Interleukin-6 as an adipokine and myokine: the regulatory role of cytokine in adipose tissue and skeletal muscle metabolism. *Hum Mov*. 2012; 13: 372---9, <http://dx.doi.org/10.2478/v10038-012-0045-y>.
47. Paula FMM, Leite NC, Vanzela EC, et al. Exercise increases pancreatic -cell viability in a model of type 1 diabetes through IL-6 signaling. *FASEB J*. 2015; 29: 1805---16, <http://dx.doi.org/10.1096/fj.14-264820>.
48. Shephard RJ, Johnson N. Effects of physical activity upon the liver. *Eur J Appl Physiol*. 2015; 115: 1-46, <http://dx.doi.org/10.1007/s00421-014-3031-6>.
49. Lustosa LP, Máximo Pereira LS, Coelho FM, et al. Impact of an exercise program on muscular and functional performance and plasma levels of interleukin 6 and soluble receptor tumor necrosis factor in prefrail community-dwelling older women: a randomized controlled trial. *Arch Phys Med Rehabil*. 2013; 94: 660-6, <http://dx.doi.org/10.1016/j.apmr.2012.11.013>.
50. He Z, Tian Y, Valenzuela PL, et al. Myokine/Adipokine Response to "Aerobic" Exercise: Is It Just a Matter of Exercise Load? *Front Physiol*. 2019; 10(4): 1379-1406. <https://doi.org/10.3389/fphys.2019.00691>
51. Pedersen BK, Febbraio MA. Muscle as an Endocrine Organ: Focus on Muscle-Derived Interleukin-6. *Physiol Rev*. 2008; 88(4): 1379-1406. <https://doi.org/10.1152/physrev.90100.2007>
52. Löffler D, Müller U, Scheuermann K, et al. Serum Irisin Levels Are Regulated by Acute Strenuous Exercise. *J Clin Endocrinol Metab*. 2015; 100(4): 1289-1299. <https://doi.org/10.1210/jc.2014-2932>
53. Broholm C., Laye M.J., Brandt C. et al. LIF is a contraction-induced myokine stimulating human myocyte proliferation. *J. Appl. Physiol*. 2011; 111(1): 251-259;
54. Otis J.S., Burkholder T.J., Pavlath G.K. Stretch-induced myoblast proliferation is dependent on the COX2 pathway. *Exp. Cell Res*. 2005; 310: 417-425;
55. Pedersen B.K., Steensberg A., Fischer C. et al. Searching for the exercise factor: is IL-6 a candidate? *J. Muscle Res. Cell. Motil*. 2003; 24(2-3): 113-119;
56. Pedersen L., Olsen C.H., Pedersen B.K. et al. Muscle-derived expression of the chemokine CXCL1 attenuates diet-induced obesity and improves fatty acid oxidation in the muscle. *Am. J. Physiol. Endocrinol. Metab*. 2012; 302(7): 831-840.
57. Shestopalov, A. V., Ganenko, L. A., Grigorieva, T. V., Laikov, A. V., Vasiliev, I. Yu., Kolesnikova, I. M., etc. Adipokines and myokines as indicators of obesity phenotypes and their relationship with indicators of intestinal microbiome diversity. *Bulletin of the Russian State Medical University*. 2023;1: 49-58. DOI: 10.24075/vrgmu.2023.004
58. Shestopalov A.V., Davydov V.V., Tumanyan G.T., Teplyakova E.D., Shkurat T.P., Mashkina E.V., Shkurat M.A., Gaponov A.M., Borisenko O.V., Roumiantsev S.A. The content of adipokines and myokines in the blood of children and adolescents with different genotypes according to the polymorphism rs662 of the paraoxonase-1 gene. *Obesity and metabolism*. 2023; 20(3): 227
59. Adipokines and myokines as indicators of obese phenotypes and their association with the gut microbiome diversity indices. *Bulletin of Russian State Medical University*. 2023; 1: 45-54
60. Kirk B, Feehan J, Lombardi G, Duque G. Curr Osteoporos Muscle, Bone, and Fat Crosstalk: The Biological Role of Myokines, Osteokines, and Adipokines. *Rep*. 2020 Aug; 18(4): 388-400. DOI: 10.1007/s11914-020-00599-y. PMID: 32529456 Review
61. Yang M., Chen P., Jin H. et al. Circulating levels of irisin in middle-aged first-degree relatives of type 2 diabetes mellitus – correlation with pancreatic β -cell function. *Diabetol. Metab. Syndr*. 2014; 6(10): 133-139;
62. Huh J.Y., Siopi A., Mougios V. et al. Irisin in response to exercise in humans with and without metabolic syndrome. *J. Clin. Endocrinol. Metab*. 2015; 100: 453-457;
63. Li Y., Li F., Lin B. et al. Myokine IL-15 regulates the crosstalk of co-cultured porcine skeletal muscle satellite cells and preadipocytes // *Mol. Biol. Rep*. 2014 V. 41, No. 11 P. 7543–7553.,
64. Pierce J.R., Maples J.M., Hickner R.C. IL-15 concentrations in skeletal muscle and subcutaneous adipose tissue in lean and obese humans: local effects of IL-15 on adipose tissue lipolysis. *Am. J. Physiol. Endocrinol. Metab*. 2015; 308(12): 1131-1139.
65. Blüher S., Panagiotou G., Petroff D. et al. Effects of a 1-year exercise and lifestyle intervention on irisin, adipokines, and inflammatory markers in obese children. *Obesity*. 2014; 22(7): 1701-1708
66. Datta N.S. Muscle-bone and fat-bone interactions in regulating bone mass: do PTH and PTHrP play any role? *Endocrine*. 2014; 47: 389-400;
67. Lai X., Price C., Lu X.L. et al. Imaging and quantifying solute transport across periosteum: implications for muscle bone crosstalk. *Bone*. 2014; 66: 82-89.
68. Henriksen T., Green C., Pedersen B.K. Myokines in myogenesis and health. *Recent Pat. Biotechnol*. 2012; 6(3): 167-171.
69. Laurens C, Bergouignan A, Moro C. Exercise-Released Myokines in the Control of Energy Metabolism. *Front Physiol*. 2020; 11: 91. <https://doi.org/10.3389/fphys.2020.00091>
70. Ahima RS, Park H-K. Connecting Myokines and Metabolism. *Endocrinol Metab*. 2015; 30(3): 235. <https://doi.org/10.3803/EnM.2015.30.3.235>
71. A systematic review of "myokines and metabolic regulation" Henry H. León-Ariza, María P. Mendoza-Navarrete, María I. Maldonado-Arango, Daniel A. Botero-Rosas October 2018 *Apunts Med Esport*. 2018; 53(200): 155-162.
72. Giudice J, Taylor JM. Muscle as a paracrine and endocrine organ. *Curr Opin Pharmacol*. 2017; 34: 49-55. <http://dx.doi.org/10.1016/j.coph.2017.05.005>
73. Barbalho SM, Flato UAP, Tofano RJ, Goulart RA, Guiguer EL, Dregiachi CRP, Buchaim DV, Araújo AC, Buchaim RL, Reina FTR, Biteli P, Reina DOBR, Bechara MD. Physical Exercise and Myokines: Relationships with Sarcopenia and Cardiovascular Complications. *Int J Mol Sci*. 2020; 21(10): 3607. PMID: 32443765. PMCID: PMC7279354.
74. Laskou F., Fuggle N.R., Patel H.P., Jameson, Cooper C., Dennison E. Associations of osteoporosis and sarcopenia with

- frailty and multimorbidity among participants of the Hertfordshire Cohort Study. *J Cachexia Sarcopenia Muscle*. 2022; 13: 220-229 <https://doi.org/10.1002/jcsm.12870>
75. Severinsen M.C.K., Pedersen B.K., Muscle-organ crosstalk: the emerging roles of myokines. *Endocr. Rev.* 2020; 41 <https://doi.org/10.1210/endrev/bnaa016>
76. Braun T, Gautel M. Transcriptional mechanisms regulating skeletal muscle differentiation, growth and homeostasis. *Nat Rev Mol Cell Biol.* 2011; 12(6): 349-361. doi: <https://doi.org/10.1038/nrm3118>
77. Han HQ, Zhou X, Mitch WE, Goldberg AL. Myostatin/activin pathway antagonism: Molecular basis and therapeutic potential. *Int J Biochem Cell Biol.* 2013; 45(10): 2333-2347. doi: <https://doi.org/10.1016/j.biocel.2013.05.019>
78. Chan MHS, Carey AL, Watt MJ, et al. Cytokine gene expression in human skeletal muscle during concentric contraction: evidence that IL-8, like IL-6, is influenced by glycogen availability. *Am J Physiol Regul Integr Comp Physiol.* 2004; 287(2): R322-7. <https://doi.org/10.1152/ajpregu.00030.2004>.
79. Heinrich PC, Behrmann I, Haan S, et al. Principles of interleukin (IL)-6-type cytokine signalling and its regulation. *Biochem J.* 2003; 374(1): 1-20. <https://doi.org/10.1042/bj20030407>.
80. Line Pedersen and Pernille Hojman Muscle-to-organ cross talk mediated by myokines *Adipocyte*. *Landes Bioscience* . 2012; 1(3): 164–167.
81. Rekha Balakrishnan, Debbie C Thurmond Mechanisms by Which Skeletal Muscle Myokines Ameliorate Insulin Resistance *Int J Mol Sci* 2022; 23(9): 4636. PMID: 35563026 PMCID: PMC9102915 DOI: 10.3390/ijms23094636.
82. Tkachuk VA, Vorotnikov AV. Molecular Mechanisms of Insulin Resistance Development. *Diabetes mellitus*. 2014; 17(2): 29-401 [In Russ.]] doi: 10.14341/DM2014229-40 <https://cyberleninka.ru/article/n/molekulyarnye-mehanizmy-razvitiya-rezistentnosti-k-insulinu>
83. Lackey DE, Olefsky JM. Regulation of metabolism by the innate immune system. *Nat Rev Endocrinol.* 2016; 12(1): 15-28. doi: 10.1038/nrendo.2015.189
84. Martyshev-Poklad A.V., Yankevich D.S., Petrova M.V., Savitskaya N.G. Two models of the development of insulin resistance and a strategy to combat age-related diseases: a review of the literature. *Problems of Endocrinology*. 2022; 68(4): 59-68. <https://doi.org/10.14341/probl13090>
85. Stafeev IS, Menshikov MY, Tsokolaeva ZI, et al. Molecular Mechanisms of Latent Inflammation in Metabolic Syndrome. Possible Role of Sirtuins and Peroxisome Proliferator-Activated Receptor Type gamma. *Biochemistry (Mosc)*. 2015; 80(10): 1217-1226. doi: 10.1134/S0006297915100028
86. Boura-Halfon S, Zick Y. Phosphorylation of IRS proteins, insulin action, and insulin resistance. *Am J Physiol Endocrinol Metab*. 2009; 296(4): E581-591. doi: 10.1152/ajpendo.90437.2008
87. Morino K, Petersen KF, Shulman GI. Molecular mechanisms of insulin resistance in humans and their potential links with mitochondrial dysfunction. *Diabetes*. 2006; 55 Suppl 2: S9-S15. doi: 10.2337/db06-S002
88. Samuel VT, Petersen KF, Shulman GI. Lipid-induced insulin resistance: unravelling the mechanism. *Lancet*. 2010; 375(9733): 2267-2277. doi: 10.1016/s0140-6736(10)60408-4
89. Liu YF, Herschkovitz A, Boura-Halfon S, et al. Serine phosphorylation proximal to its phosphotyrosine binding domain inhibits insulin receptor substrate 1 function and promotes insulin resistance. *Mol Cell Biol*. 2004; 24(21): 9668-9681. doi: 10.1128/MCB.24.21.9668-9681.2004
90. Zick Y. Uncoupling insulin signalling by serine/threonine phosphorylation: a molecular basis for insulin resistance. *Biochem Soc Trans*. 2004; 32(Pt 5): 812-816. doi: 10.1042/BST0320812
91. Hojlund K. Metabolism and insulin signaling in common metabolic disorders and inherited insulin resistance. *Dan Med J*. 2014; 61(7): B4890.
92. Stafeev IS, Vorotnikov AV, Ratner EI, et al. Latent Inflammation and Insulin Resistance in Adipose Tissue. *Int J Endocrinol*. 2017; 2017: 5076732. doi: 10.1155/2017/5076732
93. Choi JH, Banks AS, Estall JL, et al. Anti-diabetic drugs inhibit obesity-linked phosphorylation of PPARgamma by Cdk5. *Nature*. 2010; 466(7305): 451-456. doi: 10.1038/nature09291
94. Oriente F, Fernandez Diaz LC, Miele C, et al. Prep1 deficiency induces protection from diabetes and increased insulin sensitivity through a p160-mediated mechanism. *Mol Cell Biol*. 2008; 28(18): 5634-5645. doi: 10.1128/MCB.00117-08
95. Oriente F, Cabaro S, Liotti A, et al. PREP1 deficiency downregulates hepatic lipogenesis and attenuates steatohepatitis in mice. *Diabetologia*. 2013; 56(12): 2713-2722. doi: 10.1007/s00125-013-3053-3
96. Penkov DN, Egorov AD, Mozgovaya MN, Tkachuk VA. Insulin resistance and adipogenesis: role of transcription and secreted factors. *Biochemistry (Mosc)*. 2013; 78(1): 8-18. doi: 10.1134/S0006297913010021
97. Erickson HP. Irisin and FNDC5 in retrospect: An exercise hormone or a transmembrane receptor? *Adipocyte*. 2013; 2(4): 289-293. doi: 10.4161/adip.26082
98. Bostrom P, Wu J, Jedrychowski MP, et al. A PGC1-alpha-dependent myokine that drives brown-fat-like development of white fat and thermogenesis. *Nature*. 2012; 481(7382): 463-468. doi: 10.1038/nature10777
99. Chaudhury, A.; Duvoor, C.; Reddy Dendi, V.S.; Kraleti, S.; Chada, A.; Ravilla, R.; Marco, A.; Shekhawat, N.S.; Montales, M.T.; Kuriakose, K.; et al. Clinical Review of Antidiabetic Drugs: Implications for Type 2 Diabetes Mellitus Management. *Front. Endocrinol*. 2017; 8: 6.
100. Bouzakri, K.; Plomgaard, P.; Berney, T.; Donath, M.Y.; Pedersen, B.K.; Halban, P.A. Bimodal Effect on Pancreatic β -Cells of Secretory Products from Normal or Insulin-Resistant Human Skeletal Muscle. *Diabetes*. 2011; 60: 1111–1121.
101. Narendran, P.; Jackson, N.; Daley, A.; Thompson, D.; Stokes, K.; Greenfield, S.; Charlton, M.; Curran, M.; Solomon, T.; Nouwen, A.; et al. Exercise to preserve β -cell function in recent-onset Type 1 diabetes mellitus (EXTOD)—A randomized controlled pilot trial. *Diabet. Med*. 2017; 34: 1521–1531.
102. Paula, F.M.M.; Leite, N.C.; Vanzela, E.C.; Kurauti, M.A.; Freitas-Dias, R.; Carneiro, E.M.; Boscherio, A.C.; Zoppi, C.C. Exercise increases pancreatic β -cell viability in a model of type 1 diabetes through IL-6 signaling. *FASEB J*. 2015; 29: 1805–1816.
103. Camporez, J.P.G.; Jornayvaz, F.; Petersen, M.C.; Pesta, D.; Guigni, B.; Serr, J.; Zhang, D.; Kahn, M.; Samuel, V.T.; Jurczak, M.; et al. Cellular Mechanisms by Which FGF21 Improves Insulin Sensitivity in Male Mice. *Endocrinology*. 2013; 154: 3099–3109.
104. Cuevas-Ramos, D.; Aguilar-Salinas, C.A.; Gómez-Pérez, F.J. Metabolic actions of fibroblast growth factor 21. *Curr. Opin. Pediatr*. 2012; 24: 523–529.
105. Amir Levy, Y., Chiaraldi, T. P., Mudaliar, S. R., Phillips, S. A., and Henry, R. R. Excess IL-8 secretion by skeletal muscle in type 2 diabetes impairs tube growth: potential role of PI3K and Tie2 receptor. *A.M. Physiology. Endocrinol. Metabolite*. 2015; 309: E22–E34. doi: 10.1152/ajpendo.00513.2014
106. Ellingsgaard, H.; Hauselmann, I.; Schuler, B.; Habib, A.M.; Baggio, L.L.; Zeman-Meier, D.; Eppler, E.; Bouzakri, K.; Wueest, S.; Muller, Y.; et al. Interleukin-6 enhances insulin secretion by increasing glucagon-like peptide-1 secretion from L cells and alpha cells. *Nat. Med*. 2011; 17: 1481–1489.
107. Li, Z.; Yang, Y.-L.; Zhu, Y.-J.; Li, C.-G.; Tang, Y.-Z.; Ni, C.-L.; Chen, L.-M.; Niu, W.-Y. Circulating Serum Myonectin Levels in Obesity and Type 2 Diabetes Mellitus. *Exp. Clin. Endocrinol. Diabetes* 2021; 129: 528–534.
108. Barlow, J.P.; Solomon, T.P. Do skeletal muscle-secreted factors influence the function of pancreatic β -cells? *Am. J. Physiol. Metab*. 2018; 314: E297–E307.
109. Raschke, S.; Elsen, M.; Gassenhuber, H.; Sommerfeld, M.; Schwahn, U.; Brockmann, B.; Jung, R.; Wisloff, U.; Tjonna, A.E.;

- Raastad, T.; et al. Evidence against a beneficial effect of irisin in humans. *PLoS ONE*. 2013; 8: e73680.
110. Wei-Hua Jia, Nuo-Qi Wang, Lin Yin 1, Xi Chen, Bi-Yu Hou, Gui-Fen Qiang, Chi Bun Chan, Xiu-Ying Yang, Guan-Hua Du Effect of skeletal muscle phenotype and gender on fasting-induced myokine expression in mice *Biochem Biophys Res Commun*. 2019;; 514(2): 407-414. doi: 10.1016/j.bbrc.2019.04.155.
111. Erin E Terry, Xiping Zhang, Christy Hoffmann, Laura D Hughes 3, Scott A Lewis 1, Jiajia Li 1, Matthew J Wallace, Lance A Riley, Collin M Douglass, Miguel A Gutierrez-Monreal, Nicholas F Lahens, Ming C Gong, Francisco Andrade, Karyn A Esser, Michael E Hughes Transcriptional profiling reveals extraordinary diversity among skeletal muscle tissues *Elife*. 2018; 7: e34613. doi: 10.7554/eLife.34613.
112. I.I. Dedov, V.A. Tkachuk, N.B. Gusev, V.P. Shirinsky A.V. Vorotnikov, T.N. Kochegura A.Yu. Mayorov, M.V. Shestakova Type 2 diabetes mellitus and metabolic syndrome: molecular mechanisms, key signaling pathways and identification of biotargets for new drugs © Russian Association of Endocrinologists. 2018., doi: 10.14341/DM9730 *Diabetes Mellitus*. 2018; 21(5): 364-375
113. Hong-KunWu, YanZhang, Chun-MeiCao, XinliHu, MengFang, YuanYao, LiJin, GengJiaChen, PengJiang, ShuoZhang, RuishengSong, WeiPeng, FenghuaLiu, JiaojiaoGuo, LifeiTang, YanyunHe, Dan Shan, Jin Huang, Zhuan Zhou, Dao Wen Wang, Fengxiang Lv, Rui-Ping Xiao Regulates Systemic Insulin Response and Metabolic Homeostasis *Circulation*. 2019 Feb 12; 139(7): 901-914. doi: 10.1161/CIRCULATIONAHA.118.037216. PMID: 30586741 DOI: 10.1161/CIRCULATIONAHA.118.037216
114. Zhengtang Qi, Jie Xia, Xiangli Xue, Jiatong Liu, Xue Zhang, Xingtian Li, Wenbin Liu, Lu Cao, Lingxia Li, Zhiming Cui, Zhuochun Huang, Benlong Ji, Qiang Zhang, Shuzhe Ding, Weina Liu Stress-induced myonectin improves glucose homeostasis by inhibiting glycemic response to HPA axis doi: <https://doi.org/10.1101/838003>
115. Salikhova Y., Mindubayeva F., Shukurov F., Niyazova Y., Nauryzov N., Khalimova F., Bilalova D., Kharisova N., Akimzhanova N.. Comparative study of Heart Rate variability in pregnant women living in conditions of High-mountain Hypoxia of the Pamirs and the steppe zone of Central Kazakhstan. *Research Journal of Pharmacy and Technology* 2023; 16(7): 3269-4. doi: 10.52711/0974-360X.2023.00538.
116. Liu R., Zhang Q., Peng N. et al. Inverse correlation between serum irisin and cardiovascular risk factors among Chinese overweight/obese population. *BMC Cardiovasc. Disord*. 2021; 21:Art. No. 570.
117. Mai S., Grugni G., Mele C. et al. Irisin levels in genetic and essential obesity: clues for a potential dual role. *Sci. Rep*. 2020. 10: Art. No. 1020.
118. Choi Y.-K., Kim M.-K., Bae K. H. et al. Serum irisin levels in new-onset type 2 diabetes. *Diabetes Res. Clin. Pract*. 2013; 100: 96-101.
119. Zhu D., Wang H., Zhang J. et al. Irisin improves endothelial function in type 2 diabetes through reducing oxidative/nitritive stresses. *J. Mol. Cell. Cardiol*. 2015; 87: 138-147.
120. Bayrasheva V.K. Modeling of diabetes mellitus and diabetic nephropathy in experiment. *Modern problems of science and education*. 2015; 4. URL: <https://science-education.ru/ru/article/view?id=21024> (access date: 02/09/2024). UDC 616.61:616.379-008.64:612.084 DOI 10.17513/spno.127-21024
121. Bayrasheva V.K., Babenko A.Yu., Dmitriev Yu.V., Bairamov A.A., Chefu S.G., Shatalov I.S., Pehelin I.Yu., Ivanova A.N., Grineva E.N. A novel model of type 2 diabetes and diabetic nephropathy in rats. *Translational Medicine*. 2016; 3(4): 44-55. (In Russ.) <https://doi.org/10.18705/2311-4495-2016-3-4-44-55>
122. N. Zakharova, T. A. Kironenko, K. G. Milovanova, A. A. Orlova, E. Yu. Dyakova, Yu. G. Kalinnikova, A. V. Chibalin, L. V. The influence of forced running loads on the content of myokines in the skeletal muscles of mice with a model of type II diabetes mellitus 2022 *Kapilevich Russian Physiological Journal named after. THEM. Sechenova*. 2021; 107(6-7): 864-875 -10-06T22:36:55Z
123. K. G. Milovanova, I. Yu. Shuvalov, A. V. Moiseenko, L. V. Kapilevich The influence of dynamic loads on the concentration of myokines in the plasma of mice. *Physical culture, healthcare and education: materials of the XV International Scientific and Practical conference dedicated to the memory of V. S. Pirussky*. Tomsk. 2021: 345-351.
124. L. V. Kapilevich, T. A. Kironenko, A. N. Zakharova et al. Content of interleukins 6 and 15 in plasma in mice after exercise. *Receptors and intracellular signaling: international conference, May 22-25, 2017 g.: collection of articles. Pushchino*. 2017; 1: 737-742. URL: <http://vital.lib.tsu.ru/vital/access/manager/Repository/vtls:000646250>
125. L. V. Kapilevich, A. N. Zakharova, T. A. Kironenko et al. The influence of running loads on body weight and the content of myokines in skeletal muscles in diabetes mellitus (experimental study). *Innovative transformations in the field physical culture, sports and tourism: collection of materials of the XXIV All-Russian Scientific and Practical Conference. September 27 – October 2, 2021, Novomikhailovsky. Rostov n/d, 2021. pp. 220-224.* URL: <http://vital.lib.tsu.ru/vital/access/manager/Repository/koha:000894195>
126. Kironenko, K. G. Milovanova et al. Treadmill training effect on the myokines content in skeletal muscles of mice with a metabolic disorder model. *Frontiers in Psychology*. 2021; 12: 709039 (1-13). URL: <http://vital.lib.tsu.ru/vital/access/manager/Repository/koha:000892334>
127. Kironenko T. A. Production of myokines and the concentration of monovalent cations in the muscle tissue of mice during physical exercise: dissertation for the degree of candidate of biological sciences: 03.03.01 / Kironenko Tatyana Aleksandrovna; scientific hands Kapilevich L. V.; state Univ. - Tomsk: [b.i.], 2021. URL: <http://vital.lib.tsu.ru/vital/access/manager/Repository/koha:000718311>
128. Graf C, Ferrari N. Metabolic Health-The Role of AdipoMyokines. *Int J Mol Sci*. 2019; 20 (24): 6159. DOI: 10.3390/ijms20246159.
129. Doumatey AP, Bentley AR, Zhou J, Huang H, Adeyemo A, Rotimi CN. Paradoxical Hyperadiponectinemia is Associated With the Metabolically Healthy Obese (MHO) Phenotype in African Americans. *J Endocrinol Metab*. 2012; 2(2): 51–65. DOI: 10.4021/jem95W.
130. Blüher S. Metabolically healthy obesity from childhood to adulthood — does weight status alone matter? *Metabolism*. 2014; 63: 1084–109.
131. Wang HY. Plasma asprosin concentrations are increased in individuals with glucose dysregulation and correlated with insulin resistance and first-phase insulin secretion. *Mediators of Inflammation*. 2018; ID 9471583.
132. Amor M, Itariu BK, Moreno-Viedma V, Keindl M, Jürets A, Prager G, et al. Serum Myostatin is Upregulated in Obesity and Correlates with Insulin Resistance in Humans. *Exp Clin Endocrinol Diabetes*. 2019; 127 (8): 550–6.
133. Staiger H, Keuper M, Berti L, Hrabe de Angelis M, Häring HU. Fibroblast Growth Factor 21-Metabolic Role in Mice and Men. *Endocr Rev*. 2017; 38 (5): 468–88.
134. Hu C, Zhang X, Zhang N, Wei WY, Li LL, Ma ZG, et al. Osteocin attenuates inflammation, oxidative stress, apoptosis, and cardiac dysfunction in doxorubicin-induced cardiotoxicity. *Clin Transl Med*. 2020; 10 (3): e124.
135. Fu J, Li Y, Esangbedo IC, Li G, Feng D, Li L, et al. Circulating Osteonectin and Adipokine Profiles in Relation to Metabolically Healthy Obesity in Chinese Children: Findings From BCAMS. *J Am Heart Assoc*. 2018; 7 (23): e009169. DOI: 10.1161/JAHA.118.009169

- 136.Fiuza-Luces C, Garatachea N, Berger NA, Lucia A. Exercise is the real polypill. *Physiology* (Bethesda) 2013; 28(5): 330–58. doi:10.1152/physiol.00019.2013
- 137.Eckardt K, Görgens SW, Raschke S, Eckel J. Myokines in insulin resistance and type 2 diabetes. *Diabetologia*. 2014; 57(6): 1087–99. doi:10.1007/s00125-014-3224-x
- 138.Steensberg A, Febbraio MA, Osada T, Schjerling P, van Hall G, Saltin B, et al. Interleukin-6 production in contracting human skeletal muscle is influenced by pre-exercise muscle glycogen content. *J Physiol* 2001; 537(Pt 2): 633–9. doi:10.1111/j.1469-7793.2001.00633.x
- 139.Al-Khalili L, Bouzakri K, Glund S, Lönnqvist F, Koistinen HA, Krook A. Signaling specificity of interleukin-6 action on glucose and lipid metabolism in skeletal muscle. *Mol Endocrinol* (2006) 20(12): 3364–75. doi:10.1210/me.2005-0490 PubMed Abstract | CrossRef Full Text | Google Scholar
- 140.Carey AL, Steinberg GR, Macaulay SL, et al. Interleukin6 increases insulin-stimulated glucose disposal in humans and glucose uptake and fatty acid oxidation in vitro via AMP-activated protein kinase. *Diabetes*. 2006; 55: 2688–97. <http://dx.doi.org/10.2337/db05-1404>
- 141.Harder-Lauridsen NM, Krogh-Madsen R, Holst JJ, Plomgaard P, Leick L, Pedersen BK, et al. Effect of IL-6 on the insulin sensitivity in patients with type 2 diabetes. *Am J Physiol Endocrinol Metab* (2014) 306(7): E769–78. doi:10.1152/ajpendo.00571.2013 PubMed Abstract | CrossRef Full Text | Google Scholar
- 142.Jiang LQ, Duque-Guimaraes DE, Machado UF, Zierath JR, Krook A. Altered response of skeletal muscle to IL-6 in type 2 diabetic patients. *Diabetes* (2013); 62(2): 355–61. doi:10.2337/db11-1790
- 143.Prokopchuk O, Liu Y, Wang L, Wirth K, Schmidtbleicher D, Steinacker JM. Skeletal muscle IL-4, IL-4R α , IL-13 and IL-13R α 1 expression and response to strength training. *Exerc Immunol Rev* (2007); 13: 67–75.
- 144.Görgens SW, Raschke S, Hølvén KB, Jensen J, Eckardt K, Eckel J. Regulation of follistatin-like protein 1 expression and secretion in primary human skeletal muscle cells. *Arch Physiol Biochem* (2013); 119(2): 75–80. doi:10.3109/13813455.2013.768270 PubMed Abstract | CrossRef Full Text | Google Scholar
- 145.Lee HJ, Lee JO, Lee YW, Kim SA, Park SH, Kim HS. Kalirin, a GEF for Rac1, plays an important role in FSTL-1-mediated glucose uptake in skeletal muscle cells. *Cell Signal* (2016); 29: 150–7. doi:10.1016/j.cellsig.2016.10.013 CrossRef Full Text | Google Scholar
- 146.Colberg SR, Sigal RJ, Yardley JE, Riddell MC, Dunstan DW, Dempsey PC, et al. Physical activity/exercise and diabetes: a position statement of the American Diabetes Association. *Diabetes Care* (2016); 39(11): 2065–79. doi:10.2337/dc16-1728
- 147.Brian P Carson The Potential Role of Contraction-Induced Myokines in the Regulation of Metabolic Function for the Prevention and Treatment of Type 2 Diabetes 2017; May 2: 8: 97. doi: 10.3389/fendo.2017.00097. eCollection 2017. PMID: 28512448 PMCID: PMC5411437 DOI: 10.3389/fendo.2017.00097
- 148.Guo T, Jou W, Chanturiya T, et al. Myostatin Inhibition in Muscle, but Not Adipose Tissue, Decreases Fat Mass and Improves Insulin Sensitivity. *Calbet JAL, ed. PLoS One*. 2009; 4(3): e4937. <https://doi.org/10.1371/journal.pone.0004937>
- 149.Bond ND, Guo J, Hall KD, McPherron AC. Modeling Energy Dynamics in Mice with Skeletal Muscle Hypertrophy Fed High Calorie Diets. *Int J Biol Sci*. 2016; 12(5): 617–630. <https://doi.org/10.7150/ijbs.13525>
- 150.Zhang C, McFarlane C, Lokireddy S, et al. Myostatindeficient mice exhibit reduced insulin resistance through activating the AMP-activated protein kinase signalling pathway. *Diabetologia*. 2011; 54(6): 1491–1501. <https://doi.org/10.1007/s00125-011-2079-7>
- 151.Hoppeler H. Molecular networks in skeletal muscle plasticity. *J Exp Biol*. 2016;219:205–13, <http://dx.doi.org/10.1242/jeb.128207>.
- 152.Belaya ZhE, Smirnova OM, Dedov II. Role of exercise in health and in diabetes mellitus. *Problemy endokrinologii*. 2005; 51(2): 28–37. (In Russ)]]
- 153.Danilov K., Sidorenko S., Milovanova K., Klimanova E., Kapilevich L., Orlov S. Electrical pulse stimulation decreases electrochemical Na⁺ and K⁺ gradients in C2C12 myotubes. *Biochemical and Biophysical Research Communications* Volume 493, Issue 2, 18 November 2017: 875-878 <https://doi.org/10.1016/j.bbrc.2017.09.133>
- 154.Kainulainen H, Papaioannou KG, Silvennoinen M, et al. Myostatin/ activin blocking combined with exercise reconditions skeletal muscle expression profile of mdx mice. *Mol Cell Endocrinol*. 2015; 399: 131-142. <https://doi.org/10.1016/j.mce.2014.10.001>
- 155.Ko I.G., Jeong J.W., Kim Y.H., et al. Aerobic Exercise Affects Myostatin Expression in Aged Rat Skeletal Muscles: A Possibility of Antiaging Effects of Aerobic Exercise Related with Pelvic Floor Muscle and Urethral Rhabdosphincter. *Int Neurourol J*. 2014; 18(2): 77. <https://doi.org/10.5213/inj.2014.18.2.77>
- 156.MingHong Shao, QiYang Wang, QiuNan Lv, YuQiong Zhang, GuoXi Gao, Sheng Lu Advances in the research on myokine-driven regulation of bone metabolism Open Access Published: November 19, 2023. DOI:<https://doi.org/10.1016/j.heliyon.2023.e22547>
- 157.Khalafi M, Alamdari KA, Symonds ME, et al. Impact of acute exercise on immediate and following early postexercise FGF-21 concentration in adults: systematic review and meta-analysis. *Hormones*. 2021; 20(1): 23-33. <https://doi.org/10.1007/s42000-020-00245-3>
- 158.Hansen JS, Pedersen BK, Xu G, et al. Exercise-Induced Secretion of FGF21 and Follistatin Are Blocked by Pancreatic Clamp and Impaired in Type 2 Diabetes. *J Clin Endocrinol Metab*. 2016; 101(7): 2816-2825. <https://doi.org/10.1210/jc.2016-1681>
- 159.Zhang Y, Li R, Meng Y, et al. Irisin Stimulates Browning of White Adipocytes Through Mitogen-Activated Protein Kinase p38 MAP Kinase and ERK MAP Kinase Signaling. *Diabetes*. 2014; 63(2): 514-525. <https://doi.org/10.2337/db13-1106>
- 160.Xiong X-Q, Chen D, Sun H-J, et al. FNDC5 overexpression and irisin ameliorate glucose/lipid metabolic derangements and enhance lipolysis in obesity. *Biochim Biophys Acta - Mol Basis Dis*. 2015; 1852(9): 1867-1875. <https://doi.org/10.1016/j.bbdis.2015.06.017>
- 161.Miyamoto-Mikami E, Sato K, Kurihara T, et al. Endurance TrainingInduced Increase in Circulating Irisin Levels Is Associated with Reduction of Abdominal Visceral Fat in Middle-Aged and Older Adults. *Kaser S, ed. PLoS One*. 2015; 10(3): e0120354. <https://doi.org/10.1371/journal.pone.0120354>
- 162.Park M-J, Kim D-I, Choi J-H, et al. New role of irisin in hepatocytes: The protective effect of hepatic steatosis in vitro. *Cell Signal*. 2015; 27(9): 1831-1839. <https://doi.org/10.1016/j.cellsig.2015.04.010>
- 163.Sujin Kim, Ji-Young Choi, Sohee Moon, Dong-Ho Park, Hyo-Bum Kwak, Ju-Hee KangPflugers Arch . 2019; Mar; 471(3): 491-505. doi: 10.1007/s00424-019-02253-8. Epub 2019 Jan 9. Roles of myokines in exercise-induced improvement of neuropsychiatric function expand PMID: 30627775 DOI: 10.1007/s00424-019-02253-8
- 164.Pedersen BK. The disease of physical inactivity and the role of myokines in muscle-fat cross talk. *J Physiol*. 2009; 587 (Pt 23): 5559-5568. doi: 10.1113/jphysiol.2009.179515. Epub 2009 Sep 14. Review.
- 165.Kapilevich L. V., Kironenko T. A., Zakharova A. N. [et al.] The content of interleukins 6 and 15 in plasma in mice after exercise // Receptors and intracellular signaling: international conference, 22-25 May 2017: collection of articles. Pushchino, 2017; 1: 737-742. URL: <http://vital.lib.tsu.ru/vital/access/manager/Repository/vtls:000646250>
- 166.Zakharova A. N. Features of cerebral hemodynamics and the production of myokines during physical activity of various types:

- dissertation ... candidate of Biological Sciences. 2017; 171: <http://www.dslib.net/fiziologia/osobennosti-cerebralnoj-gemodinamiki-i-produkcija-miokinov-pri-fizicheskikh.html>
167. Kapilevich L. V., Kironenko T. A., Zakharova A. N. [et al.] The content of interleukins 6 and 15 in plasma in mice after exercise // Receptors and intracellular signaling: international conference, 22-25 May 2017: collection of articles. Pushchino, 2017; 1: 737-742. URL: <http://vital.lib.tsu.ru/vital/access/manager/Repository/vtls:000646250>
168. Zakharova A. N., Kironenko T. A., Milovanova K. G., Orlova A. A., Dyakova E. Yu., Kalinnikova Yu. G., A. V. Chibalin, L. V. Kapilevich Russian physiological journal named after. THEM. Sechenova, 2021, T. 107, No. 6-7: 864-875 The effect of forced running loads on the content of myokines in the skeletal muscles of mice with a model of type II diabetes mellitus Date of publication in the registry: 2022-10-06T22:36:55Z
169. Kapilevich L. V. Myokines as a promising marker of metabolic disorders and physical activity . L. Kapilevich, S. Orlov, A. Kabachkova. AIP Conference Proceedings. 2015. Vol. 1688.:030030-1-030030-4. URL: <http://vital.lib.tsu.ru/vital/access/manager/Repository/vtls:000524691>
170. Kapilevich L. V. Myokines as a promising marker of metabolic disorders and physical activity / L. Kapilevich, S. Orlov, A. Kabachkova. AIP Conference Proceedings. 2015. Vol. 1688.:030030-1-030030-4. URL: <http://vital.lib.tsu.ru/vital/access/manager/Repository/vtls:000524691>
171. Milovanova K. G., Shuvalov I. Yu., Moiseenko A. V., Kapilevich L. V. The influence of dynamic loads on the concentration of myokines in the plasma of mice. Physical culture, health care and education: materials of the XV International Scientific and Practical Conference, dedicated to the memory of V. S. Prussky, November 18, 2021 Toms, 2021.:345-351. URL: <http://vital.lib.tsu.ru/vital/access/manager/Repository/koha:000896674>
172. Milène Catoire I, Marco Mensink, Eric Kalkhoven, Patrick Schrauwen, Sander Kersten Physiological genomics. 1 Apr 2014;46(7):256-67. doi:10.1152/physiolgenomics.00174.2013. Epub 2014, February 11. Original text: Identification of human exercise-induced myokines using secretome analysis PMID: 24520153 DOI: 10.1152/physiolgenomics.00174.2013
173. Berezina A.V., Belyaeva O.D., Bazhenova E.A. and others. Features of fat oxidation during physical activity of varying intensity in patients with abdominal obesity // Probl. endocrinology. 2010. No. 56. (2). pp. 20-26
174. Zhu, Y., Liu, Q., Zhu, Y., Guo, T., Jin, M., Hao, J., Qi, C., Miao, X., Xi, D., Fan, J. and Li, J. (2022) Effects of Common Myokines on Diabetes Mellitus. Journal of Diabetes Mellitus, 12, 153-166. doi: 10.4236/jdm.2022.123013.
175. Timofeev A.V. Review of clinical trials of new methods for the prevention and treatment of diabetes mellitus in children and adolescents "Hi+Med. High technologies in medicine." 2013; No. 2 (16): 44-46.
176. <https://dni24.com/exclusive/101096-rossiyskie-uchenye-razrabatyvayut-novyy-metod-lecheniya-autizma-i-diabeta.html>
177. Gérald J Prud'homme Prevention of autoimmune diabetes by DNA vaccination Expert Rev Vaccines. 2003; Aug; 2(4): 533-40. DOI: 10.1586/14760584.2.4.533 PMID: 14711337
178. Bryan Ceballos, Michael Alexander, Jonathan R.T. Lakey Advanced Approaches in Immunotherapy for the Treatment of Type 1 Diabetes Mellitus EMJ Diabet. 2020; DOI/10.33590/emjdiabet/20-00062. <https://doi.org/10.33590/emjdiabet/20-00062>.
179. Mai Charlotte Krogh Severinsen, Bente Klarlund Pedersen Myokines in Muscle-Organ Crosstalk Endocrine Reviews, Volume 41, Issue 4, August 2020; bnaa016 Pages 594-609, <https://doi.org/10.1210/endrev/bnaa016>
180. P. Kumar, Titi Xavier Mangalathil, Vikas Choudhary. An experimental study to assess the effectiveness of structured teaching programme on knowledge regarding the management of diabetes mellitus among G.N.M. students in selected nursing school at Sikar, Rajasthan. Asian J. Management. 2014; 5(3): July-September, 329-331.
181. Chauhan H.V. In-Vivo Antidiabetic, Lipid Lowering and Antioxidant Activities of Methanolic Extract of Lawsonia inermis Leaves. Research J. Pharm. and Tech. 4(5): May 2011; Page 764-767.
182. Bhutkar M. A., Bhise S. B. Comparative Studies on Antioxidant Activity of Some Antidiabetic Plants. Research J. Pharm. and Tech. 2011; 4(9): 1409-1412.
183. Harinarayan Singh Chandel, A K Pathak, Mukul Tailang. Characterization of Some Herbal Antidiabetic Drugs in Polyherbal Formulation by Microscopy. Research J. Pharm. and Tech. 2011; 4 (1): 131-145.
184. TE Gopala Krishna Murthy, C Mayuren. Effect of Ramipril on the Pharmacodynamics of Gliclazide in Diabetic Rats. Research J. Pharm. and Tech. 2009; 2(1): Jan.-Mar. 120-122.
185. Sarika S. Lokhande, Raje V. N., More S. S., Pawar S. S. Role of Pharmacist in Prevention and Management of Diabetics. Asian Journal of Pharmaceutical Research. 2023; 13(2): 95-8 DOI: 10.52711/2231-5691.2023.00019
186. Klimontov V.V., Berikov V.B., Saik O.V. Artificial intelligence in diabetology. Diabetes mellitus. 2021; 24(2): 156-166. <https://doi.org/10.14341/DM12665>
187. Schönenberger KA, Cossu L, Prendin F, et al. Digital solutions to diagnose and manage postbariatric hypoglycemia. Front Nutr. 2022; 9. <https://doi.org/10.3389/fnut.2022.855223>
188. Suplotova L.A., Alieva O.O. Evolution of blood glucose self-monitoring technology. Diabetes mellitus. 2023; 26(6): 566-574. (In Russ.) <https://doi.org/10.14341/DM13063>
189. Prachi Gupta, Manju Bala, Sanjeev Gupta, Anita Dua, Rajesh Dabur, Elisha R. Injeti, Ashwani Mittal Efficacy and risk profile of anti-diabetic therapies: Conventional vs traditional drugs—A mechanistic revisit to understand their mode of action. Pharmacological Research. 2016; 113 (Pt A): 636-674 DOI:10.1016/j.phrs.2016.09.029
190. Fabian Sanchis-Gomar, Sergio Lopez-Lopez, Carlos Romero-Morales, Nicola Maffulli, Giuseppe Lippi, Helios Pareja-Galeano Neuromuscular Electrical Stimulation: A New Therapeutic Option for Chronic Diseases Based on Contraction-Induced Myokine Secretion REVIEW article Front. Physiol., 28 November 2019 Sec. Striated Muscle Physiology Volume 10 - 2019